**YEPD agar**

<table>
<thead>
<tr>
<th></th>
<th>1000 mL</th>
<th>500 mL</th>
<th>250 mL</th>
<th>100 mL</th>
</tr>
</thead>
<tbody>
<tr>
<td>Yeast extract</td>
<td>10 g</td>
<td>5 g</td>
<td>2.5 g</td>
<td>1 g</td>
</tr>
<tr>
<td>Peptone</td>
<td>20 g</td>
<td>10 g</td>
<td>5 g</td>
<td>2 g</td>
</tr>
<tr>
<td>Dextrose (glucose)</td>
<td>20 g</td>
<td>10 g</td>
<td>5 g</td>
<td>2 g</td>
</tr>
<tr>
<td>Agar</td>
<td>20 g</td>
<td>10 g</td>
<td>5 g</td>
<td>2 g</td>
</tr>
<tr>
<td>dH₂O</td>
<td>1000 mL</td>
<td>500 mL</td>
<td>250 mL</td>
<td>100 mL</td>
</tr>
</tbody>
</table>

1. Weigh out dry ingredients into flask that has at least twice the volume as the amount of media to be made.

2. Use a graduated cylinder to measure out the appropriate volume of ddH₂O, pour into flask.

3. Note, the agar will not dissolve, that’s OK, it will dissolve in the autoclave. Do not shake to mix so that the agar flakes do not adhere to the side of the flask.

4. Cover flask top with foil; be generous with the aluminum foil.

5. Put a small piece of autoclave tape on the foil.

6. Autoclave on the liquid setting for 20 minutes.

7. When the flask is removed from the autoclave, put in 55°C water bath for approximately 15 minutes until the agar has cooled to 55°C.

8. Add the appropriate amount of antibiotic, if using (Typically, the stock solution will be 1000X so you will add the same number of μL as there are mL of media). Swirl gently to mix antibiotic.

9. Pour plates using sterile technique.

NOTE: you can figure to use about 25-30 mL of media per plate. Usually plates are made in batches of 500 mL for easy pouring.

1000 mL = 33 to 40 plates
500 mL = 16 to 20 plates
250 mL = 8 to 10 plates
100 mL = 3 to 4 plates